Ann Clin Biochem OnlineFirst, published on March 12, 2015 as doi:10.1177/0004563215579695

DOI: 10.1177/0004563215579695

# Annals of Clinical Biochemistry

## Cystatin C and creatinine based equations in the assessment of renal function in HIV positive patients prior to commencing Highly Active Antiretroviral Therapy (HAART)

Journal:	Annals of Clinical Biochemistry
Manuscript ID:	ACB-14-258.R1
Manuscript Type:	Original Article
Date Submitted by the Author:	10-Feb-2015
Complete List of Authors:	Seape, Tebogo; University of the Witwatersrand and National Health Laboratory Service, Chemical Pathology Gounden, Verena; University of KZN, Chemical Pathology van Deventer, Hendrick; University of the Witwatersrand and National Health Laboratory Service, Chemical Pathology Candy, Geoffrey; University of Witwatersrand, Department of Surgery George, Jaya; University of Witwatersrand, Chemical Pathology; National Health Laboratory Services, Chemical Pathology
Keywords:	Creatinine < Analytes, Renal disease < Clinical studies

SCHOLARONE<sup>™</sup> Manuscripts

3	1	Cystatin C- and creatinine-based equations in the assessment of renal function in HIV positive
4 5	2	patients prior to commencing Highly Active Antiretroviral Therapy (HAART)
6		
7 8	3	Tebogo Seape <sup>1</sup> , Verena Gounden <sup>1, 2</sup> , Hendrick E van Deventer <sup>1, 3</sup> , Geoffrey P Candy <sup>4</sup> , Jaya A George <sup>1</sup>
9 10	4	<sup>1</sup> Department of Chemical Pathology, University of the Witwatersrand and National Health Laboratory
11	-	
12 13	5	Services, Charlotte Maxeke Johannesburg Academic Hospital, Parktown, Johannesburg, South Africa
14 15	6	<sup>2</sup> Current address: Department of Chemical Pathology, University of Kwa Zulu Natal and National Health
16	7	Laboratory Services Inkesi Albert Lythuli Central Hespitel Cete Manon Dunken South Africa
17	7	Laboratory Services, Inkosi Albert Luthuli Central Hospital, Cato Manor, Durban, South Africa
18		
19	8	<sup>3</sup> Lancet Laboratories, Auckland Park, Johannesburg, South Africa
20		
21	0	
22	9	<sup>4</sup> Department of Surgery, University of the Witwatersrand, Johannesburg, South Africa
23		
24	10	Competing Interests: None.
25		
26		
27	11	Funding: Siemens diagnostics supplied the kits for cystatin C determination and the National Health
28	10	
29	12	Laboratory Services funded the measured GFR determination but neither funder played any role in study
30	13	design, analysis or interpretation of data.
31	15	design, analysis of interpretation of data.
32		
33	14	Ethical approval: The Human Research Ethics Committee of the University of the Witwatersrand
34 35		
36	15	(Clearance certificate number M10410) approved the study.
37		
38	16	Guarantor: JAG
39	10	
40		
41	17	Contributorship: T S collected the data, obtained ethical permission and wrote the first draft. G C
42		
43	18	carried out the radioactive studies and contributed to the final draft, V G and H E van Deventer carried
44	19	out statistical analysis, interpretation and revisions to the article and J A George conceptualized the
45	19	out statistical analysis, interpretation and revisions to the article and J A George conceptualized the
46	20	study, interpreted data and revised the article. All authors reviewed and edited the manuscript and
47		
48	21	approved the final version of the manuscript
49		
50	22	A sheep with the second state of the second state of the second state of the state of the Nicking of The state
51	22	Acknowledgements: We thank the patients who participated in this study, the National Health
52	23	Laboratory Service for funding mGFR, and Siemens Diagnostics for the cystatin C assays.
53	25	Eutoratory betwee for funding more, and oremens Diagnosites for the cystatin c assays.
54		
55	24	Competing Interests: None
56		
57	25	Corresponding author: Jaya A George Email: jaya.george@wits.ac.za
58 50	25	Corresponding aution, Jaya A Ocorge Email. Jaya.gcorge @ wits.ac.za
59 60		
60		

#### **Annals of Clinical Biochemistry**

Page 2 of 22 DOI: 10.1177/0004563215579695

1	
2	
3	
4	
D G	
6	
1	
8	
9	
10	
11	
12	
1/	
14	
16	
17	
18	
19	
20	
21	
22	
23	
$\begin{array}{c} 2 \\ 3 \\ 4 \\ 5 \\ 6 \\ 7 \\ 8 \\ 9 \\ 1 \\ 1 \\ 1 \\ 2 \\ 1 \\ 1 \\ 1 \\ 1 \\ 1 \\ 1$	
25	
26	
27	
28	
29	
30	
31	
32	
33	
34	
35	
36	
37	
38	
39	
40	
41	
42 43	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
54	
55 56	
56	
57 50	
58 59	
60	

1	Abstract:
T	ADSTRACT:

2	
2	<b>Background:</b> We evaluated the accuracy and precision of creatinine- and cystatin C-based prediction
3	equations for estimating glomerular filtration rate (eGFR) compared to measured GFR (mGFR) in an
4	antiretroviral-naive human immunodeficiency virus (HIV) population.
5	Methods: The study population consisted of 100 treatment-naive HIV patients. GFR was estimated
6	using the Cockcroft-Gault, Modification of Diet in Renal Disease (MDRD) and Chronic Kidney Disease
7	Epidemiology Collaboration (CKD-EPI) equations as well as cystatin C-based equations (CKD-EPI
8	<sub>cystatin C</sub> , cystatin C <sub>van Deventer</sub> and CKD-EPI <sub>combined</sub> ) compared to <sup>51</sup> Cr-EDTA plasma clearance mGFR. We
9	calculated percentage bias, standard deviation (SD) of the differences, accuracy within 15% and 30% of
10	mGFR and sensitivity and specificity for predicting mGFR < $60 \text{ mL/min/1.73m}^2$ .
11	<b>Results</b> : Bias for all eGFR equations ranged from -9.4 % to 38.4 %. The CKD-EPI <sub>combined</sub> without
12	ethnicity correction factor equation had the least bias, 2.9% (-2.9 to 8.8). Bias was higher for the MDRD
13	and CKD-EPI equation with the African American ethnicity factor (38.4% and 33.7%) than without
14	(14.2% and 15.3%). SD of the differences ranged from 29.2 % (CKD-EPI combined without ethnicity
15	factor) to 54.0% (MDRD with ethnicity factor). Accuracy within 30% of mGFR ranged from 78% for
16	CKD-EPI combined without ethnicity factor to 56.7% for the Cockcroft-Gault equation. Sensitivity for
17	creatinine based equations was less than 50% and for the CKD-EPI $_{\text{cystatin C}}$ equation was 75%.
18	Conclusion: Sensitivity of creatinine-based equations for predicting GFR was poor in this group of
19	patients. The CKD-EPI combined equation performed better than creatinine-based equations.
20	
21	
21	
22	Keywords: estimated GFR, cystatin C, HIV, MDRD, prediction equations
23	Word count: 2928
24	
25	
26	
20	

1	
2	
3	INTRODUCTION
4	Human immunodeficiency virus (HIV) infection affects almost 25 million people in Sub-Saharan Africa
5	(1). While the introduction of antiretroviral therapy has resulted in a significant reduction of acquired
6	immune deficiency syndrome (AIDS) related deaths (2, 3), disease conditions such as chronic kidney
7	disease (CKD) have emerged as important causes of morbidity and mortality in the United States and
8	Europe (4). Renal dysfunction progressing to end-stage renal disease is a common complication of HIV
9	infection (5, 6). In Africa, a wide spectrum of renal diseases that have been described on biopsies
10	obtained from infected patients (7, 8). The classic kidney disease of HIV infection is HIV associated
11	nephropathy (HIVAN) (9, 10) which results in accelerated progression to AIDS and increased mortality.
12	Treatment of HIV and comorbidities may also lead to renal disease e.g. the use of tenofovir disoproxil
13	fumarate (TDF), a nucleotide reverse transcriptase inhibitor (11).
14	In addition to the effects of HIV infection and its therapy on the kidney, chronic diseases like diabetes
15	and hypertension add to the risk of CKD in Africa. The impact of both infectious diseases like HIV and
16	chronic diseases on the prevalence of CKD has huge public health implications for Africa (12). Studies
17	suggest that reduced glomerular filtration rate (GFR) is very prevalent among HIV infected people in
18	Africa (13, 14) but there is substantial variability between the methods used to estimate GFR (10, 13, 15,
19	16). In Africa renal function is most commonly assessed by either the Modification of Diet in Renal
20	Disease (MDRD) study equation (17) or the Cockcroft-Gault equation (18). More recently the creatinine
21	based CKD-EPI equation (CKD-EPI) was shown to estimate GFR (eGFR) more accurately than the
22	MDRD equation (19) and has replaced the MDRD equation for estimating GFR in some laboratories, but
23	its use has not been validated in local populations. In 2012, Inker et al, demonstrated in a large cross-
24	sectional analysis that a combined creatinine- and cystatin C-based equation (CKD-EPI combined) estimated
25	GFR more accurately than the CKD-EPI equation (20).
26	Cystatin C is a low molecular weight (13kD) non-glycosylated basic protein produced by all nucleated
27	cells (21). It is produced at a constant rate, is freely filtered by the glomerulus and does not re-enter the
28	circulation after being filtered through the glomerulus (22). It therefore meets many of the key criteria of

an ideal endogenous glomerular filtration rate marker. Cystatin C production is independent of muscle mass and dietary influences and cystatin C-based prediction equations is therefore potentially not subject to some of the limitations of serum creatinine-based eGFR equations (23). A previous study from our centre showed that cystatin C-based prediction equations are more precise than serum creatinine-based equations for patients in predicting eGFR in patients with measured GFR (mGFR) > 60 ml/min/1.73m<sup>2</sup> (24). Cystatin C may therefore be of benefit in detection of early renal dysfunction. As patients with HIV are at increased risk for the development of CKD, it is important that eGFR equations can accurately estimate GFR and identify patients with possible CKD.

9 The aim of this study was to evaluate the use of commonly described eGFR prediction equations in an
 10 antiretroviral naive HIV positive population using <sup>51</sup>Cr-EDTA plasma clearance as the reference mGFR.

#### 12 Materials and Methods:

Sample collection: This cross-sectional study was conducted on samples collected from one hundred treatment naïve HIV positive adult medical inpatients at the Chris Hani Baragwanath Hospital, Soweto, South Africa. Exclusion criteria were: patients receiving HAART (currently and those who defaulted treatment), pregnant or breastfeeding individuals and patients with one or more of the following conditions hypertension, diabetes mellitus, oedema or presence of known renal complications with current admission. The Human Research Ethics Committee of the University of the Witwatersrand (Clearance certificate number M10410) approved the study. We obtained written informed consent from all patients prior to enrolment. Participants' clinical history, age, height and gender were available from medical records.

## 22 Patients were fasting from the night before. Five mL of EDTA plasma and five mL of serum was

23 collected from each participant for cystatin C and creatinine measurement respectively between 8:00 am

- 24 and 10 am. Samples were centrifuged at 3500 rpm for 10 minutes and stored at  $-70^{\circ}$ C until analysis.
- 25 GFR Measurement: GFR measurement was performed by the Nuclear Medicine department using <sup>51</sup>Cr-
- 26 EDTA according to published guidelines (25). After injection of 3.7 MBq <sup>51</sup>Cr-EDTA intravenously
- 27 blood samples were collected from the contralateral arm at 120 and 240 minutes post injection. GFR was
- 28 measured with the slope intercept method. The Brochner-Mortensen equation was used according to

1	guidelines adopted by the British Nuclear Medicine Society to correct the obtained measurements (26).
2	The DuBois method BSA (m <sup>2</sup> ) = $[71.84 \text{ weight (kg)}^{0.425} * \text{height (cm)}^{0.725}] / 10000 \text{ was used to}$
3	normalize GFR to the body surface area (27).
4	Creatinine and cystatin C measurement: Cystatin C was measured on a Siemens Advia 1800 analyser
5	(Siemens Healthcare Diagnostics, Tarrytown, USA) by an automated latex-enhanced immune-
6	nephelometric assay traceable to the IFCC reference method (ERM-DA471). Creatinine was measured
7	spectrophotometrically using the kinetic modified Jaffe method on the Siemens Advia 1800 analyser
8	(Siemens Healthcare Diagnostics, Tarrytown). The creatinine assay is traceable to an isotope dilution
9	mass spectrometry reference creatinine method (28). All samples were analysed in duplicate.
10	GFR estimation: Glomerular filtration rate was estimated using the Cockcroft-Gault equation (18)
11	normalized to 1.73m <sup>2</sup> , MDRD equation (IDMS traceable) with (29) and without ethnicity factor (30), the
12	CKD-EPI equation (19), the cystatin C $_{van Deventer}$ equation (24), the CKD-EPI $_{cystatin C}$ equation and the
13	CKD-EPI combined equation (20). (Refer to Supplementary material for full equations used)
14	Statistical Analysis
15	The Shapiro Wilk test was used to assess normality of data. Continuous data variables are expressed as
16	mean ±SD if parametric and median (interquartile range, IQR) if non-parametric. Difference plots were
17	used for comparison studies. For the difference plots mean percentage bias, SD of the difference and
18	95% limits of agreement were calculated for each of the equations. Wilcoxon match pairs signed ranks
19	test was used to test for significance of bias compared to reference mGFR. Accuracy within 15% $(P_{15})$
20	and within 30% ( $P_{30}$ ) was also calculated for each of the equations. The McNemar test was used to
21	compare $P_{15}$ and $P_{30}$ values. Receiver operating characteristic (ROC) curve analysis was used to calculate
22	the sensitivity and specificity to correctly predict $mGFR < 60 mL/min/1.73m^2$ . Statistical analysis was
23	performed using the MedCalc Statistical program (MedCalc Version 11.6.1, Mariakerke, Belgium) and
24	Analyse-It (Analyse-It Software Ltd Version 2.26, Leeds UK).
25	Results
26	One hundred black South African HIV positive patients were enrolled in the study. Results of ninety-
27	seven patients are presented. Three patient results were excluded due to incomplete data. All participants

28 were inpatients admitted at the Chris Hani Baragwanath hospital for a wide range of diseases except for

ა ⊿	
4	
5 6	
7	
8	
9	
10	
11	
12	
13	
14	
15	
16	
17	
18	
19	
20 21	
21	
22	
23 24	
24	
25	
26	
27	
28	
29	
30 21	
31	
32	
33 34	
35 36	
30 37	
38	
39 40	
40	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
54	
55	
56	
57	
58	
59	
60	

primary renal disease. The majority (43/97, 44%) of the patients were admitted with lower respiratory
 tract infection, followed by tuberculosis in 25% (24/97) and opportunistic infections such as cryptococcal
 meningitis in 14% (14/97). Minority of patients had diagnoses that included lymphomas and bacterial
 and viral meningitis.

Basic characteristics of the group are presented in Table 1. mGFR ranged between 24.7 and 164.6 and
mL/min/1.73m<sup>2</sup>. Of note 16% (n=16) of patients had an mGFR of less than 60 ml/min/1.73m<sup>2</sup> and 50%
of the group had a mGFR greater than 90 ml/min/1.73m<sup>2</sup>.

8

#### 9 Comparisons of the different eGFR equations:

#### 10 Bias and difference plots

11 Most of the creatinine based equations evaluated overestimated mGFR in this population. Inclusion of 12 the ethnicity factor resulted in a greater overestimation of mGFR as compared to when the ethnicity 13 factor was excluded (Table 2 and Figure 1). The MDRD equation overestimation was 38.4 (27.5 to 49.3) 14 % when the ethnicity factor was included vs. 14.2 (5.2 to 23.2) % without ethnicity factor. The CKD-15 EPI equation overestimation was 33.7 (25.0 to 42.4) % with the ethnicity factor vs. 15.3 (7.8 to 22.8) % 16 without. For the CKD-EPI combined equation overestimation was 11.5 (5.4 to 17.6) % with the ethnicity 17 factor vs. 2.9 (-2.9 to 8.8) % without the ethnicity factor. Using the Wilcoxon match pairs signed ranks 18 test, only the CKD-EPI combined without ethnicity factor equation had no significant bias when compared 19 to mGFR. The cystatin C based equations (CKD-EPI existin C and cystatin C van Deventer) underestimated GFR in this population, -5.5 (-11.7 to 0.7) % and -9.4 (-15.5 to -3.2) % respectively. 20

SD of the differences appeared to be higher for creatinine based equations compared to cystatin C based
equations (mean SD of creatinine based equations was 39.1% compared to 29.9% for cystatin C based
equations). The cystatin C-based equations (CKD-EPI <sub>combined</sub>, CKD-EPI <sub>cystatin C</sub> and cystatin C <sub>van Deventer</sub>)
showed a concentration bias effect, i.e. more positively biased at lower mGFR. There was no
concentration bias effect for the MDRD or Cockcroft-Gault equation.

26

27

1		
2	1	
3 4	T	
5 6	2	Accuracy within 15% and 30% of mGFR
7 8 9	3	Accuracy within 30% of mGFR ranged from 56.7% for the Cockcroft-Gault equation to 78.0% for the
10 11	4	CKD-EPI combined without ethnicity factor equation (Table 2). Using the McNemar test accuracy within
12 13	5	30% was higher for the MDRD equation without ethnicity factor (59.8%) than for the MDRD equation
14 15	6	with ethnicity factor, 48.3% (P=0.001). $P_{15}$ and $P_{30}$ was significantly better for the CKD-EPI equation
16	7	without the ethnicity factor (35.1% and 62.9%) compared to the CKD-EPI equation with ethnicity factor,
17 18	8	24.7% and 41.2% (P=0.05 and P<0.0001 respectively). Using the McNemar test $P_{15}$ and $P_{30}$ was not
19 20	9	significantly different between the MDRD equation without ethnicity factor and the CKD-EPI equation
21 22	10	without ethnicity factor. $P_{15}$ for the CKD-EPI <sub>cystatin C</sub> equation (42.3%) was better than for the MDRD
23 24	11	equation without ethnicity factor, 27.8% (P=0.007) but not statistically better than the CKD-EPI equation
25 26	12	without ethnicity factor (35.1%). $P_{15}$ and $P_{30}$ were significantly better for the CKD-EPI <sub>combined</sub> without
27 28	13	ethnicity factor equation than for the CKD-EPI equation without ethnicity factor (P=0.003 and P=0.002
29 30	14	respectively) (Table 2 and Addendum Table 1).
31 32	15	
33 34	16	Sensitivity for predicting mGFR < 60 mL/min/1.73m <sup>2</sup>
35 36		
37	17	While none of the creatinine bases equations had a good sensitivity for predicting $mGFR < 60$
38 39	18	mL/min/1.73m <sup>2</sup> (Table 2), sensitivity was higher for cystatin C based equations. Sensitivity for the CKD-
40 41	19	$EPI_{cystatin C}$ equation was 75% and for the CKD-EPI <sub>combined</sub> equation without ethnicity factor it was 69%.
42 43	20	For all creatinine based equations sensitivity was less than 50%.
44 45	21	
46 47	22	
48 49	22	Discussion
50 51	23	Here we report on the performance of GFR estimating equations using creatinine and cystatin C with and
52 53	24	without the ethnicity factor in a group of anti-retroviral naive HIV positive patients. The results of this
54	25	study show that eGFR varies depending on the equation used and that cystatin C based equations
55 56	26	perform better than creatinine based equations. This is in keeping with a number of other studies that
57 58 59	27	have shown cystatin C to be superior to creatinine only based equations in the general population (24,
59		-

Page 8 of 22

31) as well as in HIV infected individuals (32, 33). These results have implications for the clinical use of GFR estimating equations in the HIV positive population.

In the context of HIV infection, impaired kidney function develops as a result of various risk factors including aging, genetic factors and the burden of HIV infection itself. The World Health Organization recommends screening for renal disease using eGFR prior to initiation of therapy with TDF (34). The most commonly used equations are the MDRD and the Cockcroft-Gault equations, which result in overestimation of mGFR. Following a review of the current literature, the authors could not find recommendations as to which eGFR equation should be used for patients initiating TDF therapy although all guidelines recommend that GFR should be estimated at the beginning of therapy and periodically thereafter.

The MDRD equations has been shown to have varying accuracy in different population groups (30, 35, 36). This has been attributed to variations in non-GFR determinants of serum creatinine such as muscle mass and diet which may be affected by acute and chronic disease (37). In our study the CKD-EPI equation was not superior to the MDRD equation in antiretroviral naive HIV positive patients. This is in contrast to Inker et al. who noted that the CKD-EPI equation performed better than the MDRD equation in HIV infected patients on antiretroviral therapy (20). They used iohexol clearance as a gold standard on clinic patients while we used <sup>51</sup>Cr-EDTA plasma clearance and our patients were in patients. A systematic review of estimating equations for GFR showed that neither the MDRD nor the CKD-EPI is optimal for all population groups but that the use of the CKD-EPI would lead to a smaller average bias in clinical practice (38). It remains to be determined if this is true in African populations. In this study, including the African American ethnicity factor in the MDRD and CKD-EPI equations the resulted in an overestimation of mGFR which improved when the ethnicity factor was not included. An explanation for this may be the patient population selected, who are HIV infected hospital in-patients and

thus more likely to have decreased muscle mass due to muscle wasting and malnutrition (39, 40). The

MDRD ethnicity factor was based on an African American population (largely descendant from West Africa) with CKD and hence is applicability is likely not universal to all African populations and those

without CKD.

Wyatt et al have previously investigated estimation of GFR in an ARV naïve population, using iohexol clearance for measured GFR (36). Accuracy within 30% of mGFR was 83% when the ethnicity factor was not used compared to 73% with the race coefficient (36). This is higher than the accuracy shown in the present study. It is not clear if the alkaline picrate method they used was IDMS traceable. They used dried blood spot measurement of iohexol clearance as a gold standard on clinic patients while we used <sup>51</sup>Cr-EDTA plasma clearance. Madala et al. have also shown, using (99m)Tc-DTPA-measured GFR, that inclusion of the African-American ethnicity correction factor in black South Africans resulted in 17.1% overestimation of mGFR compared to 5% without the use of the ethnicity factor (41).

Cystatin C concentration is considered independent of muscle mass and dietary influences which may explain the better performance in our population subgroup. Of note, the mean BMI of the group was 20.9 kg/m<sup>2</sup>. While cystatin C is independent of muscle mass, it may be influenced by adiposity and by inflammation (42). This may explain the different performance of cystatin C based equations observed in a number of studies. In transplant patients, cystatin C based equations gave better 30% and 50% accuracy compared with creatinine based equations (43). Only a few studies have looked at the performance of cystatin C based equations in HIV patients often with contradictory results. In HIV patients on treatment, Inker et al showed that cystatin C was less accurate for  $GFR < 60 \text{ ml/min}/1.73 \text{m}^2$ (44), while in a small group of HIV positive patients the cystatin C van Deventer equation was more precise than the MDRD or the CKD-EPI equation (24). Similarly, cystatin C eGFR was more precise than MDRD in a group of Thai HIV patients (45). Driver et al showed that cystatin C eGFR were more strongly associated with mortality risk than creatinine eGFR (33).

There are limitations to our study, one being the relatively small sample size. As a result, we had very few patients within each CKD group and most of the study cohort did not have CKD. However although most patients in the study did not have GFR < 60 ml/min/ $1.73m^2$ , a study by Jose et al. examining the decline of renal function following initiation of TDF demonstrated that an eGFR of < 75 mL/minute/ $1.73m^2$ m<sup>2</sup> at the start of therapy was associated with an increased risk of discontinuing TDF and an eGFR of < 90 mL/min/ $1.73m^2$  at the time of discontinuation was associated with an increased risk of incomplete reversibility (46). Jose et al advise that renal monitoring during TDF therapy and discontinuation for

#### Annals of Clinical Biochemistry

those with declining renal function (46), hence the importance of using as accurate as possible means of estimating GFR in this group. We did not measure urinary protein excretion. Furthermore, the study was conducted in inpatients although GFR measurements were not carried out when they were acutely ill. Strengths of the study are that GFR was measured using a <sup>51</sup>Cr-EDTA plasma clearance method and that the study was able to evaluate various commonly used eGFR equations (both creatinine based and cystatin C based) in a population (African HIV positive antiretroviral naïve patients) for whom accurate eGFR measurement is important.

Epidemiological studies have shown that the prevalence of stage 3 CKD is about 10% in the western world (47, 48). Whilst population based data on the burden of CKD in Africa is lacking, a systematic review and meta-analysis of 21 medium and high quality studies from Africa noted a prevalence of 13.9% (95%CI 12.1-15.7) (49). In this study, using mGFR, 16% of all patients had GFR < 60 mL/min/1.73  $m^2$ . The estimated prevalence varies depending on the method used to estimate GFR (50). Among the elderly and in the general population the prevalence was shown to be much higher when the MDRD formula was used compared to when cystatin C based equations were used (51, 52). We also showed that that eGFR in our HIV infected population differed based on the eGFR equation used. Moving from the MDRD equation to the CKD-EPI combined will decrease the estimated prevalence of CKD, which has major public health implications (53). In our study, cystatin C based prediction equations had a smaller bias compared to creatinine based equations with the smallest bias was observed for the CKD-EPI combined equation. Given that cost is a prohibitive factor in Africa it may not be practical at this stage to recommend the widespread use of cystatin C and we were unable to identify a subgroup that may benefit from it. In the South African context the use of CKD-EPI or MDRD equations without ethnicity factor may be the most practical option for estimating GFR.

59 60

2 3 4	1	J	References
5 6	2	1.	UNAIDS. The Joint United Nations Programme on HIV/AIDS. UNAIDS/WHO "AIDS Epidemic
7 8	3		Update: December 2007. 2007.
9 10	4	2.	Mocroft A, Vella S, Benfield TL, Chiesi A, Miller V, Gargalianos P, et al. Changing patterns of
11	5		mortality across Europe in patients infected with HIV-1. EuroSIDA Study Group. Lancet.
12 13	6		1998;352(9142):1725-30.
14 15	7	3.	Vittinghoff E, Scheer S, O'Malley P, Colfax G, Holmberg SD, Buchbinder SP. Combination
16 17	8		antiretroviral therapy and recent declines in AIDS incidence and mortality. J Infect Dis.
18 19	9		1999;179(3):717-20.
20 21	10	4.	Selik RM, Byers RH, Jr., Dworkin MS. Trends in diseases reported on U.S. death certificates that
22	11		mentioned HIV infection, 1987-1999. J Acquir Immune Defic Syndr. 2002;29:378-87.
23 24	12	5.	Winston JA, Burns GC, Klotman PE. The human immunodeficiency virus (HIV) epidemic and HIV-
25 26	13		associated nephropathy. Semin Nephrol. 1998;18:373-7.
27 28	14	6.	Choi AI, Rodriguez RA, Bacchetti P, Bertenthal D, Volberding PA, O'Hare AM. The impact of HIV on
29 30	15		chronic kidney disease outcomes. Kidney Int. 2007;72(11):1380-7.
31 32	16	7.	Gerntholtz TE, Goetsch SJ, Katz I. HIV-related nephropathy: a South African perspective. Kidney Int.
33 34	17		2006;69(10):1885-91.
35	18	8.	Wearne N, Swanepoel CR, Boulle A, Duffield MS, Rayner BL. The spectrum of renal histologies seen
36 37	19		in HIV with outcomes, prognostic indicators and clinical correlations. Nephrol Dial Transplant.
38 39	20		2012;27:4109-18.
40 41	21	9.	Schwartz EJ, Szczech LA, Ross MJ, Klotman ME, Winston JA, Klotman PE. Highly active
42 43	22		antiretroviral therapy and the epidemic of HIV+ end-stage renal disease. J Am Soc Nephrol.
44 45	23		2005;16:2412-20.
46 47	24	10.	Peters PJ, Moore DM, Mermin J, Brooks JT, Downing R, Were W, et al. Antiretroviral therapy
48	25		improves renal function among HIV-infected Ugandans. Kidney Int. 2008;74(7):925-9.
49 50	26	11.	Verhelst D, Monge M, Meynard JL, Fouqueray B, Mougenot B, Girard PM, et al. Fanconi syndrome
51 52	27		and renal failure induced by tenofovir: a first case report. Am J Kidney Dis. 2002;40:1331-3.
53 54	28	12.	Wools-Kaloustian KK, Gupta SK. Will there be an epidemic of HIV-related chronic kidney disease in
55 56	29		sub-Saharan Africa? Too soon to tell. Kidney Int. 2008;74:845-7.
57 58			
50			

1	13.	Sumaili EK, Cohen EP, Zinga CV, Krzesinski JM, Pakasa NM, Nseka NM. High prevalence of
2		undiagnosed chronic kidney disease among at-risk population in Kinshasa, the Democratic Republic of
3		Congo. BMC Nephrol. 2009;10:18.
4	14.	Banda J, Mweemba A, Siziya S, Mweene M, Andrews B, Lakhi S. Prevalence and Factors Associated
5		with Renal Dysfunction in HIV Positive and Negative Adults at the University Teaching Hospital, in
6		Lusaka. Med J Zambia. 2010;37(3):136-42.
7	15.	Wools-Kaloustian K, Gupta SK, Muloma E, Owino-Ong'or W, Sidle J, Aubrey RW, et al. Renal
8		disease in an antiretroviral-naive HIV-infected outpatient population in Western Kenya. Nephrol Dial
9		Transplant. 2007;22:2208-12.
10	16.	Mulenga LB, Kruse G, Lakhi S, Cantrell RA, Reid SE, Zulu I, et al. Baseline renal insufficiency and
11		risk of death among HIV-infected adults on antiretroviral therapy in Lusaka, Zambia. AIDS.
12		2008;22(14):1821-7.
13	17.	Levey AS, Bosch JP, Lewis JB, Greene T, Rogers N, Roth D. A more accurate method to estimate
14		glomerular filtration rate from serum creatinine: a new prediction equation. Modification of Diet in
15		Renal Disease Study Group. Ann Intern Med. 1999;130(6):461-70.
16	18.	Cockcroft DW, Gault MH. Prediction of creatinine clearance from serum creatinine. Nephron.
17		1976;16(1):31-41.
18	19.	Levey AS, Stevens LA, Schmid CH, Zhang Y, Castro IIIAF, Feldman HI, et al. A New Equation to
19		Estimate Glomerular Filtration Rate. Ann Intern Med. 2009;150:604-12.
20	20.	Inker LA, Schmid CH, Tighiouart H, Eckfeldt JH, Feldman HI, Greene T, et al. Estimating glomerular
21		filtration rate from serum creatinine and cystatin C. N Engl J Med. 2012;367(1):20-9.
22	21.	Chapman HA, Jr., Reilly JJ, Jr., Yee R, Grubb A. Identification of cystatin C, a cysteine proteinase
23		inhibitor, as a major secretory product of human alveolar macrophages in vitro. Am Rev Respir Dis.
24		1990;141(3):698-705.
25	22.	Grubb A, Simonsen O, Sturfelt G, Truedsson L, Thysell H. Serum concentration of cystatin C, factor D
26		and beta 2-microglobulin as a measure of glomerular filtration rate. Acta Med Scand. 1985;218(5):499-
27		503.
28	23.	Filler G, Bokenkamp A, Hofmann W, Le Bricon T, Martinez-Bru C, Grubb A. Cystatin C as a marker
29		of GFRhistory, indications, and future research. Clin Biochem. 2005;38(1):1-8.

#### **Annals of Clinical Biochemistry**

60

13

2			
3	1	24.	van Deventer HE, Paiker JE, Katz IJ, George JA. A comparison of cystatin C- and creatinine-based
4 5	2		prediction equations for the estimation of glomerular filtration rate in black South Africans. Nephrol
6 7	3		Dialy Transplant. 2011;26:1553-8.
8 9	4	25.	Fleming JS, Zivanovic MA, Blake GM, Burniston M, Cosgriff PS, British Nuclear Medicine S.
10 11	5		Guidelines for the measurement of glomerular filtration rate using plasma sampling. Nucl Med
12	6		Commun. 2004;25(8):759-69.
13 14	7	26.	Brochner-Mortensen J. A simple method for the determination of glomerular filtration rate. Scand J
15 16	8		Clin Lab Invest. 1972;30(3):271-4.
17 18	9	27.	Du Bois D, Du Bois EF. A formula to estimate the approximate surface area if height and weight be
19 20	10		known. Arch Intern Med 1916;17:863-71.
21 22	11	28.	Myers GL, Miller WG, Coresh J, Fleming J, Greenberg N, Greene T, et al. Recommendations for
23	12		improving serum creatinine measurement: a report from the Laboratory Working Group of the National
24 25	13		Kidney Disease Education Program. Clin Chem. 2006;52:5-18.
26 27	14	29.	Levey AS, Coresh J, Greene T, Marsh J, Stevens LA, Kusek JW, et al. Expressing the Modification of
28 29	15		Diet in Renal Disease Study equation for estimating glomerular filtration rate with standardized serum
30 31	16		creatinine values. Clin Chem. 2007;53(4):766-72.
32 33	17	30.	van Deventer HE, George JA, Paiker JE, Becker PJ, Katz IJ. Estimating glomerular filtration rate in
34	18		black South Africans by use of the modification of diet in renal disease and Cockcroft-Gault equations.
35 36	19		Clin Chem. 2008;54:1197-202.
37 38	20	31.	Guo X, Qin Y, Zheng K, Gong M, Wu J, Shou W, et al. Improved glomerular filtration rate estimation
39 40	21		using new equations combined with standardized cystatin C and creatinine in Chinese adult chronic
41 42	22		kidney disease patients. Clin Biochem. 2014;47(13-14):1220-6.
43 44	23	32.	Praditpornsilpa K, Avihingsanon A, Chaiwatanarat T, Chaiyahong P, Wongsabut J, Ubolyam S, et al.
45	24		Comparisons between validated estimated glomerular filtration rate equations and isotopic glomerular
46 47	25		filtration rate in HIV patients. AIDS. 2012;26(14):1781-8.
48 49	26	33.	Driver TH, Scherzer R, Peralta CA, Tien PC, Estrella MM, Parikh CR, et al. Comparisons of creatinine
50 51	27		and cystatin C for detection of kidney disease and prediction of all-cause mortality in HIV-infected
52 53	28		women. AIDS. 2013;27:2291-9.
54 55	29	34.	WHO. Consolidated guidelines on the use of antiretroviral drugs for treating and preventing HIV
56	30		infection. 2013.
57 58			
59 60			12

1	35.	Poggio ED, Wang X, Greene T, Van Lente F, Hall PM. Performance of the modification of diet in
2		renal disease and Cockcroft-Gault equations in the estimation of GFR in health and in chronic kidney
3		disease. J Am Soc Nephrol. 2005;16:459-66.
4	36.	Wyatt CM, Schwartz GJ, Owino Ong'or W, Abuya J, Abraham AG, Mboku C, et al. Estimating kidney
5		function in HIV-infected adults in Kenya: comparison to a direct measure of glomerular filtration rate
6		by iohexol clearance. PloS one. 2013:e69601.
7	37.	Stevens LA, Coresh J, Greene T, Levey AS. Assessing kidney functionmeasured and estimated
8		glomerular filtration rate. N Engl J Med. 2006;354:2473-83.
9	38.	Earley A, Miskulin D, Lamb EJ, Levey AS, Uhlig K. Estimating equations for glomerular filtration rate
10		in the era of creatinine standardization: a systematic review. Ann Intern Med. 2012;156(11):785-95,
11		W-270, W-1, W-2, W-3, W-4, W-5, W-6, W-7, W-8.
12	39.	Salomon J, De TP, Melchior JC. Nutrition and HIV infection. Br J Nutr. 2002;87 Suppl 1:S111-9.
13	40.	Andrade CS, Jesus RP, Andrade TB, Oliveira NS, Nabity SA, Ribeiro GS. Prevalence and
14		characteristics associated with malnutrition at hospitalization among patients with acquired
15		immunodeficiency syndrome in Brazil. PloS one. 2012;7:e48717.
16	41.	Madala ND, Nkwanyana N, Dubula T, Naiker IP. Predictive performance of eGFR equations in South
17		Africans of African and Indian ancestry compared with (9)(9)mTc-DTPA imaging. Int Urol Nephrol.
18		2012;44(3):847-55.
19	42.	Stevens LA, Schmid CH, Greene T, Li L, Beck GJ, Joffe MM, et al. Factors other than glomerular
20		filtration rate affect serum cystatin C levels. Kidney Int. 2009;75:652-60.
21	43.	Harman G, Akbari A, Hiremath S, White CA, Ramsay T, Kokolo MB, et al. Accuracy of cystatin C-
22		based estimates of glomerular filtration rate in kidney transplant recipients: a systematic review.
23		Nephrol Dial Transplant. 2013;28(3):741-57.
24	44.	Inker LA, Wyatt C, Creamer R, Hellinger J, Hotta M, Leppo M, et al. Performance of creatinine and
25		cystatin C GFR estimating equations in an HIV-positive population on antiretrovirals. J Acquir
26		Immune Defic Syndr. 2012;61(3):302-9.
27	45.	Praditpornsilpa K, Avihingsanon A, Chaiwatanarat T, Chaiyahong P, Wongsabut J, Ubolyam S, et al.
28		Comparisons between validated estimated glomerular filtration rate equations and isotopic glomerular
29		filtration rate in HIV patients. AIDS. 2012;26:1781-8.

#### **Annals of Clinical Biochemistry**

0				
2 3	1	46.	Jose S, Hamzah L, Campbell LJ, Hill T, Fisher M, Leen C, et al. Incomplete reversibility of estimat	ted
4 5	2		glomerular filtration rate decline following tenofovir disoproxil fumarate exposure. J Infect Dis.	
6 7	3		2014;210(3):363-73.	
8 9	4	47.	Coresh J, Selvin E, Stevens LA, Manzi J, Kusek JW, Eggers P, et al. Prevalence of chronic kidney	
10 11	5		disease in the United States. JAMA. 2007;298(17):2038-47.	
12	6	48.	McCullough K, Sharma P, Ali T, Khan I, Smith WC, MacLeod A, et al. Measuring the population	
13 14	7		burden of chronic kidney disease: a systematic literature review of the estimated prevalence of	
15 16	8		impaired kidney function. Nephrol Dial Transplant. 2012;27(5):1812-21.	
17 18	9	49.	Stanifer JW, Jing B, Tolan S, Helmke N, Mukerjee R, Naicker S, et al. The epidemiology of chronic	с
19 20	10		kidney disease in sub-Saharan Africa: a systematic review and meta-analysis. Lancet Glob Health.	
21 22	11		2014;2:e174-e81.	
23 24	12	50.	Delanaye P, Cohen EP. Formula-based estimates of the GFR: equations variable and uncertain.	
25	13		Nephron Clin Pract. 2008;110(1):c48-53; discussion c4.	
26 27	14	51.	Delanaye P, Cavalier E, Saint-Remy A, Lutteri L, Krzesinski JM. Discrepancies between creatinine	<b>;-</b>
28 29	15		based and cystatin C-based equations in estimating prevalence of stage 3 chronic kidney disease in	an
30 31	16		elderly population. Scand J Clin Lab Invest. 2009;69(3):344-9.	
32 33	17	52.	Delanaye P, Cavalier E, Moranne O, Lutteri L, Krzesinski JM, Bruyere O. Creatinine-or cystatin C-	-
34 35	18		based equations to estimate glomerular filtration in the general population: impact on the epidemiol	logy
36 37	19		of chronic kidney disease. BMC Nephrol. 2013;14:57.	
38 39	20	53.	Delanaye P, Cavalier E, Mariat C, Maillard N, Krzesinski JM. MDRD or CKD-EPI study equations	s for
40	21		estimating prevalence of stage 3 CKD in epidemiological studies: which difference? Is this differen	ice
41 42	22		relevant? BMC Nephrol. 2010;11:8.	
43 44	23			
45 46	25			
47 48	24			
49 50	25			
51 52				
53	26			
54 55	27			
56 57	• •			
58 59	28			
60				15

# Page 16 of 22 DOI: 10.1177/0004563215579695

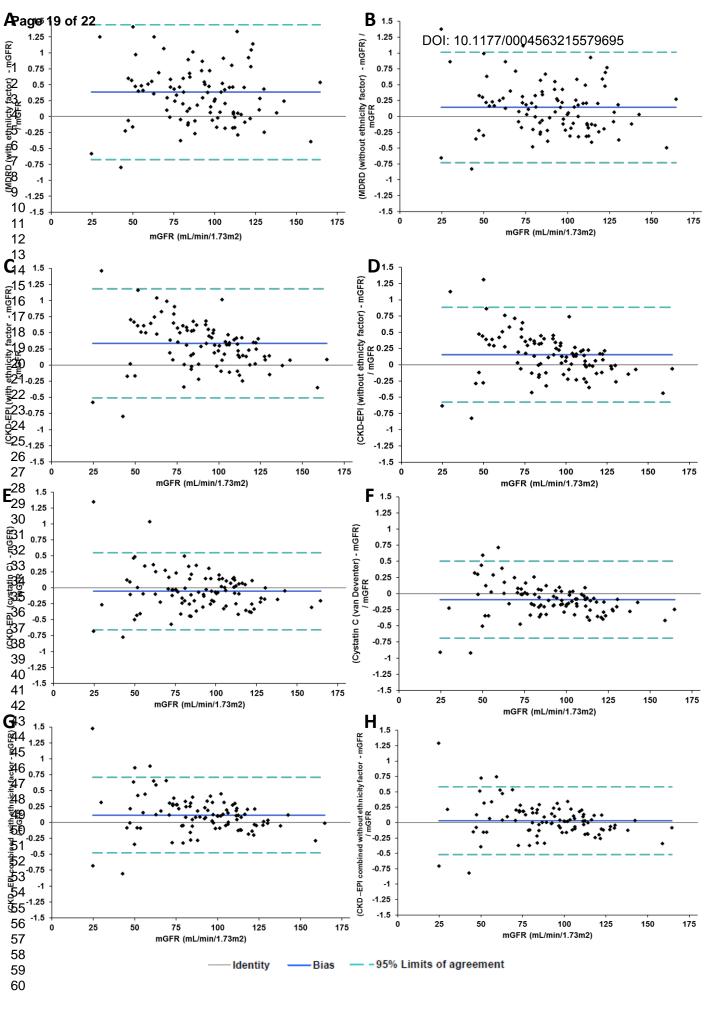
1		
2		
3	1	Legend: Figure 1
4		
5	r	Difference plots: A: MDRD with ethnicity factor B: MDRD without ethnicity factor. C: CKD-EPI with
6	2	Difference piols: A: MDRD with elimicity factor <b>B</b> : MDRD without elimicity factor. C: CKD-EFI with
7	3	ethnicity factor. D: CKD-EPI without ethnicity factor. E: CKD-EPI cystatin C F: van Deventer cystatin C G:
8	5	ennicity factor. D: CKD-EPT without ennicity factor. E: CKD-EPT <sub>cystatin C</sub> F: Vali Deventer <sub>cystatin C</sub> G:
9	4	CKD-EPI combined with ethnicity factor H: CKD-EPI combined without ethnicity factor
10	4	CRD-Li I combined with cullificity factor <b>H</b> . CRD-Li I combined without cullificity factor
11		
12	5	
13		
14		
15	6	
16		
17	7	
18	7	
19		
20	8	
20	Ũ	
22		
23	9	
23 24		
24 25	4.0	
26	10	
20		
28	11	
	11	
29		
30		
31		
32		
33		
34		
35		
36		
37		
38		
39		
40		
41		
42		
43		
44		
45		
46		
47		
48		
49		
50		
51		
52		
53		
54		
55		
56		
57		
58		
59		
60		

#### **Table 1: Participant Characteristics**

	Male (N=57)	Female (N=40)	Combined (N=97)
	Mean ± SD or	Mean ± SD or	Mean ± SD or
	Median (IQR)	Median (IQR)	Median (IQR)
Age, years	39.6 ± 9.2	33.2 ± 9.0	37.0 ± 9.6
Weight, kg	58.7 (12.4)	56.8 (25.5)	57.5 (14.4)
Height, cm	169 (9.9)	157 (6.8)	165.0 (12.3)
Body surface area, m <sup>2</sup>	1.69 ± 0.15	1.62 ± 0.18	$1.66 \pm 0.16$
Body mass index (BMI), kg/m <sup>2</sup>	20.2 (3.5)	20.1 (9.7)	20.9 (5.1)
S-creatinine, μmol/L	87.0 (38.0)	60.8 (21.1)	72.5 (43.3)
S-cystatin C, mg/L	1.0 (0.4)	1.0 (0.4)	1.0 (0.4)
CD4, / uL	152 (256)	185 (282)	162 (280)
mGFR, mL/min/1.73m <sup>2</sup>	88.1 (37.0)	95.4 (38.4)	92.5 (38.3)
Classification: mGFR	N (%)	N (%)	N (%)
mGFR < 60 ml/min/1.73m <sup>2</sup>	9 (16%)	7 (18%)	16 (16%)
mGFR 60-90 ml/min/1.73m <sup>2</sup>	21 (37%)	10 (25%)	31 (32%)
mGFR > 90 ml/min/1.73m <sup>2</sup>	27 (47%)	23 (58%)	50 (52%)
Classification: CD4	N (%)	N (%)	N (%)
≥ 500 / uL	32 (57%)	20 (53%)	52 (55%)
200 – 499 / uL	18 (32%)	17 (45%)	35 (37%)
< 200 / uL	6 (11%)	1 (3%)	7 (7.4%)

	Bias (%)	Median difference,	P <sup>1</sup>	SD of	95% Limit of	•	0		
	(95% CI)	mL/min/1.73m2 (95% CI)	Ρ	difference	Agreement	<b>P</b> <sub>15</sub>	P <sub>30</sub>	Sensitivity (95% CI)	Specificity (95% CI)
Cockcroft-Gault	22.1 (12.7 to 31.5)	14.5 (7.8 to 21.4)	< 0.0001	46.7%	-69.3% to 113.6%	34.0	56.7	0.38 (0.15 to 0.65)	0.96 (0.90 to 0.99)
MDRD – with ethnicity factor	38.4 (27.5 to 49.3)	28.2 (20.5 to 36.7)	< 0.0001	54.0%	-67.4% to 144.3%	22.7	43.3	0.31 (0.11 to 0.59)	0.99 (0.93 to 1.00)
MDRD – without ethnicity factor	14.2 (5.2 to 23.2)	15.0 (3.5 to 25.9)	0.01	44.6%	-73.1% to 101.6%	27.8	59.8	0.44 (0.20 to 0.70)	0.95 (0.88 to 0.99)
CKD-EPI – with ethnicity factor	33.7 (25.0 to 42.4)	26.7 (20.8 to 32.0)	< 0.0001	43.2%	-51.1% to 118.4%	24.7	41.2	0.31 (0.11 to 0.59)	0.99 (0.93to 1.00)
CKD-EPI – without ethnicity factor	15.3 (7.8 to 22.8)	10.2 (5.2 to 15.4)	< 0.0001	37.3%	-57.8% to 88.4%	35.1	62.9	0.31 (0.11 to 0.59)	0.98 (0.91 to 1.00)
CKD-EPI cystatin C	-5.5 (-11.7 to 0.7)	-6.6 (-11.2 to -2.1)	0.005	30.9%	-66.1% to 55.1%	42.3	75.3	0.75 (0.48 to 0.93)	0.90 (0.82 to 0.96)
Cystatin C <sub>van Deventer</sub>	-9.4 (-15.5 to -3.2)	-11.3 (-15.2 to -7.3)	< 0.0001	30.4%	-68.9% to 50.2%	45.4	72.2	0.63 (0.35 to 0.85)	0.93 (0.85 to 0.97)
CKD-EPI <sub>combined</sub> – vith ethnicity factor	11.5 (5.4 to 17.6)	8.4 (4.4 to 12.7)	< 0.0001	30.3%	-47.8% to 70.9%	44.0	73.0	0.56 (0.30 to 0.80)	0.96 (0.90 to 0.99)
CKD-EPI <sub>combined</sub> – without ethnicity factor	2.9% (-2.9 to 8.8)	-0.7 (-4.5 to 3.4)	0.52	29.2%	-54.3% to 60.2%	51.0	78.0	0.69 (0.41 to 0.89)	0.95 (0.88 to 0.99)

<sup>1</sup>Wilcoxon match pairs signed ranks test compared to mGFR



Annals of Clinical Biochemistry

Downloaded from acb.sagepub.com by guest on March 13, 2015

## Supplementary material

## a) Cockcroft-Gault equation:

eGFR = (140 - age) x body weight (kg)/a x serum creatinine in µmol/L x 1.73 m<sup>2</sup>/BSA

where: a = 0.8 for men and 0.85 for women and BSA is body surface area

## b) MDRD equation:

eGFR = 175 x (serum creatinine in  $\mu$ mol/L/88.4)<sup>-1.154</sup> x (age)<sup>-0.203</sup> x (0.742 if female) x 1.212 if African-American

## c) CKD-EPI equation (2009):

eGFR = 141 x min  $(S_{cr} / \kappa, 1)^{\alpha} x \max(S_{cr} / \kappa, 1)^{-1.209} x 0.993^{Age} x 1.018$  [if female] x 1.159 [if black]

where:  $S_{cr}$  is serum creatinine in µmol/L,  $\kappa$  is 61.9 for females and 79.6 for males,  $\alpha$  is -0.329 for females and -0.411 for males,

## d) CKD-EPI <sub>cystatin C</sub> equation (2012):

eGFR = 133 x min(SCysC/0.8, 1)<sup>0.499</sup> max(SCysC/0.8, 1)<sup>1.328</sup> x 0.996<sup>Age</sup> [ x 0.932 if female]

where: SCysC is serum cystatin C (in mg/L)

## e) Cystatin C van Deventer eGFR equation:

 $eGFR = 10^{2.35} \times 10^{(SCysC \times -0.33)} \times 10^{(-0.003 \times Age)}$ 

where: SCysC is serum cystatin C (in mg/L)

## f) CKD-EPI combined equation (2012):

eGFR =  $135 \text{ x} \min (\text{SCr/k}, 1)^a \text{ x} \max(\text{SCr/k}, 1)^{-0.601} \text{ x} \min(\text{SCysC}/0.8, 1)^{-0.375} \text{ x} \max(\text{SCysC}/0.8, 1)^{-0.711} \text{ x} 0.995^{\text{Age}} [\text{ x} 0.969 \text{ if female}] [\text{ x} 1.08 \text{ if black}]$ 

where: SCr is serum creatinine (in mg/dL), SCysC is serum cystatin C (in mg/L), k is 0.7 for

females and 0.9 for males, a is 0.248 for females and 0.207 for males

Addendum: Table 1: P value for McNemar test; Comparing P<sub>30</sub> and P<sub>15</sub> respectively

	Cockcroft-Gault	MDRD – with ethnicity factor	MDRD – without ethnicity factor	CKD-EPI – with ethnicity factor	CKD-EPI – without ethnicity factor	CKD-EPI <sub>cystatin</sub> C	Cystatin C van Deventer	CKD-EPI combined – with ethnicity factor	CKD-EPI combined – without ethnicity factor
Cockcroft-Gault		0.0009	0.65	0.003	0.29	0.02	0.004	0.001	< 0.0001
		and 0.08	and 0.26	and 0.12	and 1.00	and 0.14	and 0.31	and 0.09	and 0.005
MDRD – with			0.001	0.58	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001 and
ethnicity factor			and 0.61	and 0.21	and 0.04	and 0.005	and 0.01	and 0.002	< 0.0001
MDRD – without				0.005	0.69	0.08	0.02	0.005	0.0008
ethnicity factor				and 0.74	and 0.21	and 0.008	and 0.03	and 0.74	and <0.0001
CKD-EPI – with					< 0.0001	< 0.0001 and	< 0.0001	< 0.0001	< 0.0001
ethnicity factor					and 0.05	0.007	and 0.02	and 0.0008	and < 0.0001
CKD-EPI – without						0.18	0.06	0.02	0.002
ethnicity factor						and 0.14	and 0.37	and 0.10	and 0.003
CKD-EPI cystatin C							0.45	0.69	0.10
CKD-EFT cystatin C							and 0.63	and 1.00	and 0.28
Cystatin C van Deventer								1.00	0.36
Cystatin C van Deventer								and 0.79	and 0.15
CKD-EPI <sub>combined</sub> - with ethnicity									0.12
factor									and 0.19
CKD-EPI <sub>combined</sub> – without ethnicity factor									